(FILE 'HOME' ENTERED AT 11:04:30 ON 21 DEC 2006) FILE 'CA' ENTERED AT 11:04:38 ON 21 DEC 2006 L1 207053 S (KINASE OR (COVALENT? OR CHEMICAL?) (2A) MODIF? OR PHOSPHORYLA?) (7A) (PEPTIDE OR PROTEIN OR SUBSTRATE OR HCG OR CHORIONIC GONADOTROPHIN) 7001 S L1 AND (FLUORESC? OR LUMINESC?) L_2 79443 S (BILAB? OR DILAB? OR MULTILAB? OR (DUAL OR BI OR TWO OR 2 OR L3PLURAL? OR MULTI? OR DOUBLE) (3A) (DYE OR LABEL? OR TAG OR INDICATOR OR FLUOROPHORE OR LUMINOPHORE OR CHROMOPHORE)) 27127 S (DYE OR LABEL? OR TAG OR INDICATOR OR NON(1A) (LUMINESC? OR L4FLUORESC?) OR NONLUMINESC? OR NONFLUORESC?) (5A) (DIMER OR STACK? OR AGGREGAT? OR COMPLEX?) L5 336 S L2 AND L3 50 S L2 AND L4 L6 L7 361 S L2 AND OUENCH? 22 S L5-6 AND L7 L8L9 7 S L2 AND (HCG OR GONADOTROPHIN) L10 69 S L3 AND (HCG OR GONADOTROPHIN) L11 44 S L4 AND (HCG OR GONADOTROPHIN) L12 129 S L6, L8-11 AND PY<2000 FILE 'BIOSIS' ENTERED AT 11:14:36 ON 21 DEC 2006 L13 FILE 'MEDLINE' ENTERED AT 11:15:20 ON 21 DEC 2006 92 S L12 L14 FILE 'CA, BIOSIS, MEDLINE' ENTERED AT 11:16:44 ON 21 DEC 2006 L15 196 DUP REM L8 L12 L13 L14 (144 DUPLICATES REMOVED) => d bib, ab l15 1-196 L15 ANSWER 3 OF 196 CA COPYRIGHT 2006 ACS on STN ΑN 142:193980 CA TIUse of chimeric fluorescent proteins as reagents to determine kinase and phosphatase activity IN Violin, Jonathan D.; Newton, Alexandra C.; Tsien, Roger Y.; Zhang, Jin PASO U.S. Pat. Appl. Publ., 68 pp., Cont.-in-part of U.S. Ser. No. 865,291. PΙ US 2005026234 **A**1 20050203 US 2004-857622 20040528 US 5981200 Α 19991109 US 1997-792553 19970131 PRAI US 1996-594575 A2 19960131 AB The present invention relates generally to reagents for detq. kinase and phosphatase activity, and more specifically to chimeric proteins contq. two fluorescent proteins and a phosphorylatable domain, and methods of using such chimeric proteins to detect kinase or phosphatase activity. The present invention provides compns. and methods that are generally useful for nondestructively detecting and monitoring protein kinase and phosphatase activities in individual living eukaryotic cells, including

mammalian cells, and provide a means to obtain spatial and temporal resoln. on the order of a few micrometers and seconds, or better. disclosed herein, **protein kinase** and phosphatase activities can be

monitored using chimeric substrates (chimeric phosphorylation

indicators) that incorporate reporter mols. such as fluorescent proteins or luminescent complexes, whose properties change significantly as a function of the phosphorylation state of the substrate. Protein interactions are detected by resonance energy transfer using fluorescent proteins or lanthanide complexes to label the putative partners. compns. of the invention are adaptable to modification using methods such as high throughput combinatorial generation and screening techniques and, therefore, readily can be varied to allow monitoring of any desired kinase, phosphatase, or protein interaction. Accordingly, the present invention provides chimeric phosphorylation indicators having various structures as disclosed herein. In a first embodiment, a chimeric phosphorylation indicator contains, in operative linkage, a donor mol., a phosphorylatable domain, a phosphoaminoacid binding domain, and an acceptor mol., wherein the phosphoaminoacid binding domain specifically binds to a phosphoaminoacid when present in the phosphorylatable domain, the donor mol. and the acceptor mol. exhibit a detectable resonance energy transfer when the donor is excited, and the phosphorylatable domain and phosphoaminoacid binding domain do not substantially emit light to excite the acceptor. The authors generated genetically-encoded fluorescent reporters for PKC activity that reversibly respond to stimuli activating PKC. Specifically, phosphorylation of the reporter expressed in mammalian cells causes changes in fluorescence resonance energy transfer (FRET), allowing real time imaging of phosphorylation resulting from PKC activation. A reporter for PKC-mediated phosphorylation (C Kinase Activity Reporter, CKAR) was constructed analogously to a previously described reporter (AKAR) for protein kinase A (Zhang et al., 2001). CKAR was successfully designed to be a specific, reversible reporter of phosphorylation by PKC (FIGS. 8 and 9). The foregoing results demonstrate that a genetically engineered reporter may be used for monitoring the activity of endogenous PKC. These indicators may have significant value in flow cytometry, complex tissues, or high-throughput screening, where quant. monitoring of translocation of a GFP-tagged C1 domain, PKC, or PH domain to the plasma membrane is difficult.

- L15 ANSWER 23 OF 196 CA COPYRIGHT 2006 ACS on STN
- AN 131:99074 CA
- TI Analysis of Src Kinase and Protein Kinase C Activity by Capillary Electrophoresis and Laser-Induced Fluorescence
- AU Wu, W. -S.; Tsai, J. -L.
- CS Poison Control and Analysis Center, and Graduate Institute of Occupational Safety and Health, Kaohsiung Medical College, Kaohsiung, 80708, Taiwan
- SO Analytical Biochemistry (1999), 269(2), 423-425
- AB A sensitive capillary electrophoresis (CE)-based assay for protein kinases using peptide substrates labeled with the amino-reactive fluorescent cyanine dye, Cy5, was developed. Sensitivity was improved using a laser-induced fluorescence (LIF) detector. The assay was applied for the anal. of protein kinase C and Src kinase.
- L15 ANSWER 46 OF 196 CA COPYRIGHT 2006 ACS on STN
- AN 126:327015 CA
- TI Conformation of the N-terminal segment of a monocysteine mutant of

- troponin I from cardiac muscle
- AU Dong, Wen-Ji; Chandra, Murali; Xing, Jun; Solaro, R. John; Cheung, Herbert C.
- CS Department of Biochemistry and Molecular Genetics, University of Alabama at Birmingham, Birmingham, AL, 35294-2041, USA
- SO Biochemistry (1997), 36(22), 6745-6753
- A mono-Cys mutant of cardiac muscle troponin I, cTnI(S5C/C81I/C98S), was AB generated from a mouse cTnI cDNA clone and expressed in a bacterial system. Cys-5 was modified with the fluorescent SH group reagent, IAANS, to probe the conformation of the N-terminal extension of the mutant and the mutant complexed with cardiac muscle troponin C. emphasis was on the effect of phosphorylation of Ser-23 and Ser-24 by protein kinase A on the conformation of the N-terminal segment. Phosphorylation resulted in an 8-nm red shift of the emission spectrum of the attached IAANS probe and a redn. of its quantum yield by a factor The intensity decay of nonphosphorylated IAANS-labeled mutant of 4-5. was complex and had to be described by a sum of 3 exponential terms, with lifetimes in the range 0.1-5 ns. A 4th component in the range of 7-9 ns was required to describe the intensity decay of the phosphorylated mutant. Phosphorylation also reduced the weighted mean lifetime, consistent with the changes obsd. in the steady-state fluorescence parameters and a 33% decrease in the global rotational correlation time calcd. from anisotropy decay data. This change in correlation time suggested a decrease in the axial ratio of the protein. The fluorescence changes of the labeled mutant induced by phosphorylation were carried over to its complex with troponin C. Stern-Volmer plots of acrylamide quenching of the steady-state fluorescence were essentially linear for nonphosphorylated mutant, but displayed pronounced concave downward curvatures for the phosphorylated protein under all conditions studied. The present results were interpreted in terms of a more compact hydrodynamic shape of the phosphorylated cTnI mutant and were consistent with a folded conformation of the N-terminal extension induced by phosphorylation of the 2 Ser residues. These conformational changes may play a role in the modulation of cardiac muscle contractility by troponin I phosphorylation.
- L15 ANSWER 75 OF 196 BIOSIS on STN
- AN 1995:81915 BIOSIS
- Detection of neutral endopeptidase (NEP, enkephalinase, EC 3.4.24.11) in relation to dopaminergic and gonadoliberinergic nerve endings in the median eminence of the male rat: A double labeling ultrastructural study.
- AU Vandenbulcke, Franck; Ciofi, Philippe; Beauvillain, Jean Claude [Reprint author]
- CS INSERM U. 156, 1 place de Verdun, 59045 Lille Cedex, France
- SO Journal of Neuroendocrinology, (1994) Vol. 6, No. 6, pp. 655-664.
- The existence of neutral endopeptidase (Enkephalinase, NEP, E.C.3.4.24.11) in membranes of nerve endings in the rat median eminence suggests that some neuropeptides have paracrine and/or autocrine actions in this region. In vitro, neutral endopeptidase is capable of hydrolysing a variety of regulatory peptides but in vivo, many works indicate that in the central nervous system this enzyme is highly

implicated in the biological inactivation of enkephalins and tachykinins. In addition there is evidence that NEP is also involved in the inactivation of neurotensin in vivo. The modulation of the release of gonadotrophin releasing hormone (GnRH) is one of the documented actions of enkephalins within the median eminence. However, it is at present unclear whether enkephalins act on dopamine endings, on GnRH endings or on both. As the technical parameters and particularly the tissue fixation used to detect neutral endopeptidase are compatible with immunocytochemical detection of GnRH and tyrosine-hydroxylase (the rate limiting enzyme in the synthesis of catecholamines), two double immunolabelings were realized at the ultrastructural level to determine if GnRH and dopamine nerve endings have the enzyme inserted within their plasma membrane. Our study shows the presence of neutral endopeptidase on tyrosine-hydroxylase-immunoreactive nerve endings while presence of the enzyme on GnRH-immunoreactive nerve endings is not demonstrated. Consequently, our results provide morphological arguments for possibilities of paracrine and/or autocrine actions by neuropeptides inactivated by neutral endopeptidase on tuberoinfundibular dopaminergic nerve endings. Conversely, action of the same peptides on GnRH boutons seems more unlikely.

- L15 ANSWER 168 OF 196 CA COPYRIGHT 2006 ACS on STN
- 99:154314 CA AN
- ΤI Interaction of creatine kinase from monkey brain with substrate: analysis of kinetics and fluorescence polarization
- Grossman, Steven H. ΑU
- CS Dep. Chem., Univ. South Florida, Tampa, FL, 33620, USA
- SO
- Journal of Neurochemistry (1983), 41(3), 729-36 Titrimetric detn. of the dissocn. consts. for the binding of substrates AB to creatine kinase (I) from monkey brain reveals 13- and 4-fold synergism in the forward and reverse directions, resp. This synergism is expressed as a decrease in the KD for a given substrate in the ternary complex compared with the binary complex, and may be a reflection of substrate-induced conformational change. I labeled with 2 mols. of 5'-iodoacetamidofluorescein displays a blue shift and a decrease in fluorescence intensity on binding of MgADP, indicative of movement of the dye into a more hydrophobic environment and quenching of the extrinsic fluorescence. Rotational relaxation times detd. from anal. of fluorescence polarization of dansylated brain I decrease from 212 to 189 ns on MgADP binding. Dansylated I in 0.5% SDS has a rotational relaxation time of 135 ns. The rotational relaxation time of dansylated muscle-type isoenzyme is unaffected by MgADP and has the same value as the brain isoenzyme-MgADP complex. Polarization values at 25° for muscle and brain enzyme labeled with 3-(4-maleimidylphenyl)-7diethylamino-4-methylcoumarin compared with limiting polarization and polarization of the free dye suggest that dye rotation is severely restricted in the muscle form, but that the dye possesses freedom of rotation in the brain form. These results suggest that, compared with the muscle isoenzyme, the brain isoenzyme is more open at the active site and more flexible overall. Binding of MgADP by brain I produces a protein more compact across 1 or both of its rotational axes, thus resembling the conformation of the muscle isoenzyme. I in brain, unlike that from muscle, is probably subject to kinetic regulation accompanied

by conformational modification, suggesting that the neurobiochem. role of the brain isoenzyme is distinct from the metabolic function of the muscle isoenzyme.

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